



Sample preparation and chromatographic techniques in NADES extract analysis

Dr. Erica G. Wilson, Dr. Young Hae Choi
Leiden University, The Netherlands

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Before starting.....

- Why NADES?
- It is a different, novel option to evaluate.
- It has proved to increase solubility of “difficult” molecules.
- It does not have to be removed before use if made with use-compatible ingredients.
- Most ingredients are GRAS compliant

Not an alternative..... a welcome option



Before starting.....

Still to be done: evaluate “greenness” ...

Among others:

- LCA (life-cycle assessment): many individual ingredients have been assessed but not the NADES themselves.
 - Can they be recycled: evaluate each application to see how they rate in a circular economy framework.
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General approach for extract analysis

Qualitative information:

- Is this extract really authentic, i.e., is it made with the correct species, part and extraction solvent, method, etc.?

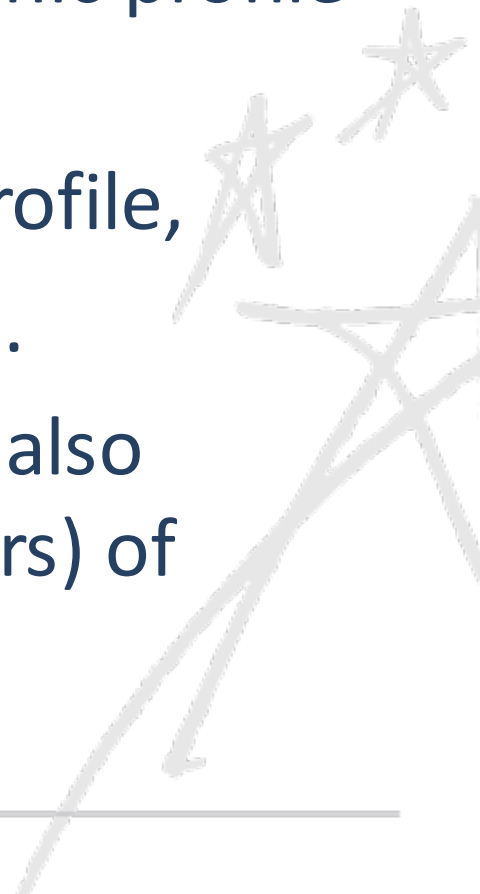
Quantitative information:

- Was the extract made with the correct ratio of material: solvent?
 - Was it made with quality starting materials?
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Qualitative information

Chemical profile

- Generally analyse the chromatographic profile such as TLC, HPLC
 - Look for coincidence of the whole profile, especially ratios of some components.
 - Presence of markers of material but also absence of markers (negative markers) of adulterants.
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NADES extracts....

Potential food /cosmetic ingredients?

- Removal of extraction solvent not required
 - Increased stability of components
 - Possible added value from extraction solvent component, e.g. xylitol
 - Concentrated source of active components
 - GRAS ingredients
 - Liquid easily combined into liquid / semi- solid formulations.
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Organic/aqueous solvent extract - NADES extract

Conventional	NADES
Low viscosity	Medium to high viscosity
Low Mr (50-100 Da range)	Relatively higher (100-360 Da)
Non-ionisable	Generally ionisable
Volatile	Non-volatile
Thermally stable	Generally thermally unstable
No acid/base character	Often acid/bases



Analytical methods

Spectroscopic:

- UV/Vis
- MS
- ^1H NMR
- IR

Chromatographic:

- HPLC/DAD
- HPLC/ELSD/RI
- LCMS
- GCMS





Sample preparation: 3 approaches

- Dilution
- Extraction
- SPE





-Sample pre-treatment (I): Dilution/injection

Applicable if:

- Samples require no pre-concentration
- No solvent incompatibility with the analytical method.

HPLC/DAD; UV/Vis

How is it done?

- Simple dilution with water or mobile phase (preferable)
 - Filtration with membrane filter (0.2 μ).
-





Sample pre-treatment (II)

Liquid-liquid extraction

Sample + water

+ Immiscible solvent

Analyte/s of interest

- Take to dryness
- Redissolve
- Inject
- Inject directly



Sample pre-treatment (III)

Solid Phase extraction (SPE)

- Dilution
- Extraction on C18 (sample retention)

or

IEX (solvent retention for ionisable
NADES)





Example: Calendula



Calendula NADES extract

(xylitol: citric acid: water) or (glucose : citric acid : water).

- Carotenoids: lutein, α - and β - carotene, etc
- Flavonoid glycosides: rutin, quercetrin, etc
- Flavonoid aglycones: quercetin, isorhamnetin, etc.

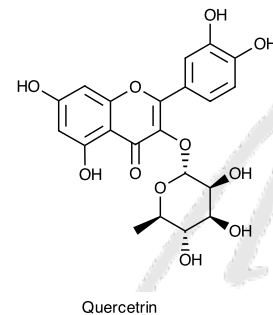
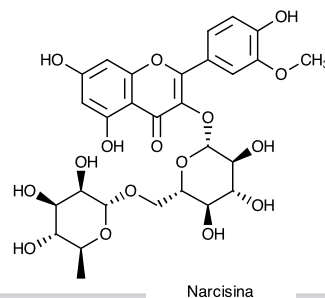
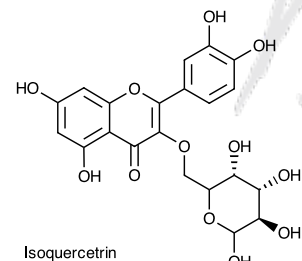
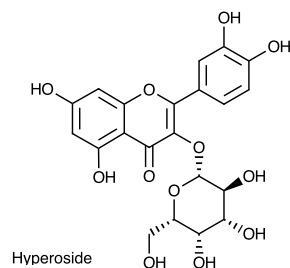
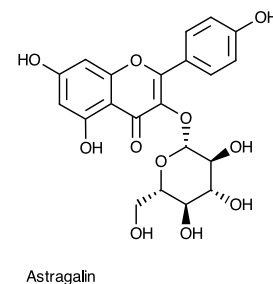
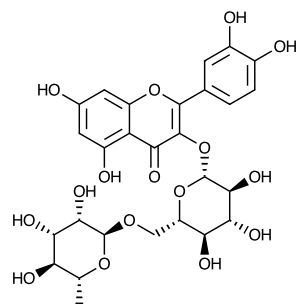
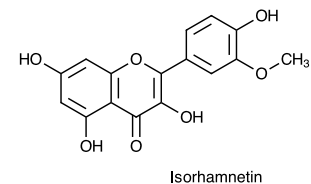
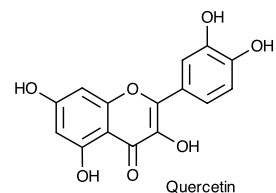




Calendula: chemical composition (I)

- Flavonoid aglycones
- Flavonoid glycosides

Hydrophilic, soluble
in methanol/water;
methanol; ethyl acetate;
butanol, etc.



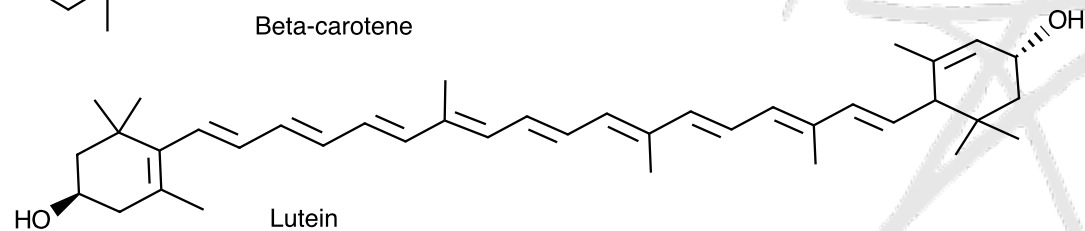
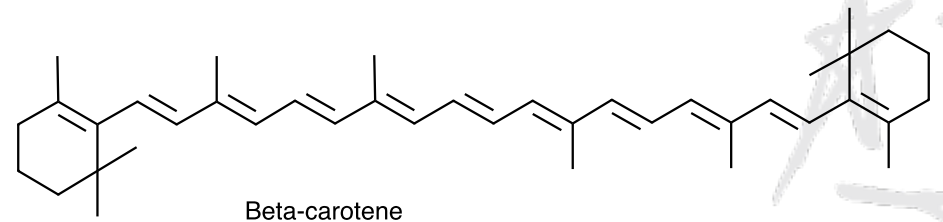
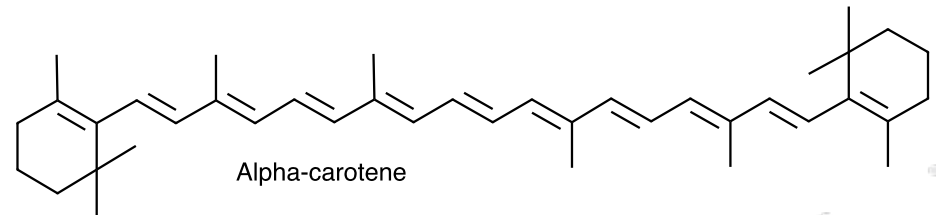


Calendula: chemical composition (II)

Carotenoids:

Lipophilic, soluble
in non-polar solvents
 CH_2Cl_2 , toluene,
hexane.

Note: in this case, water
content is critical: even
starting with fresh material
instead of dried material
reduced the yield
dramatically





Strategy for flavonoids

(i) HPLC/DAD ; UV/Vis



Direct dilution with water

(ii) LCMS; NMR



(i) Ethyl acetate

(ii) Take to dryness

(iii) Redissolve in mobile phase/
NMR solvent





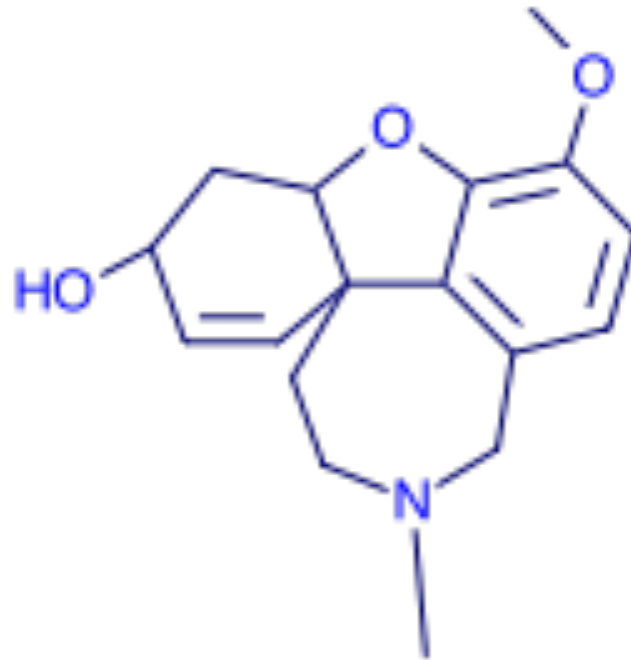
Strategy for carotenoids

For all analytical platforms:

- (i) Dilute with H_2O
- (ii) Extract with CH_2Cl_2
- (iii) Dilute with mobile phase if pre-concentration not required
- (iv) HPLC: C18 / ACN: H_2O – ethyl acetate: MeOH
or
- (v) Total carotenoid content (UV/Vis)



Example: Alkaloids in Narcissus bulb



Galanthamine

Extraction/analysis flow-chart

200 mg liophyllized bulb material
+
NADES components (solid)
+
Sand



Discharge of NADES extract
(5 min)

GCMS

Column: DB5 (30mx0.25mmx
0.25μ)

HPLC/DAD

Column: Gemini NX-C18
3.0μ C18, 100 x 4.6 mm
Mobile phase:
Methanol:water:TEA
(50:50:0.3)
Detection: 279 nm
Flow: 0.7 ml/min

GC/MS

Pre-treatment:
Basic extraction
(NaOH→Cl₃CH)
Take to dryness
Redissolve with MeOH
Derivatise

HPLC/DAD

Pre-treatment:
Sample+MeOH (2 h)
Dilution with mobile/phase
Centrifuge
Adjust pH≈11
Filter with PVDF/RC 0.2μ




NADES alkaloid extract

Issues to consider:

Can a NADES component interact with analytes?

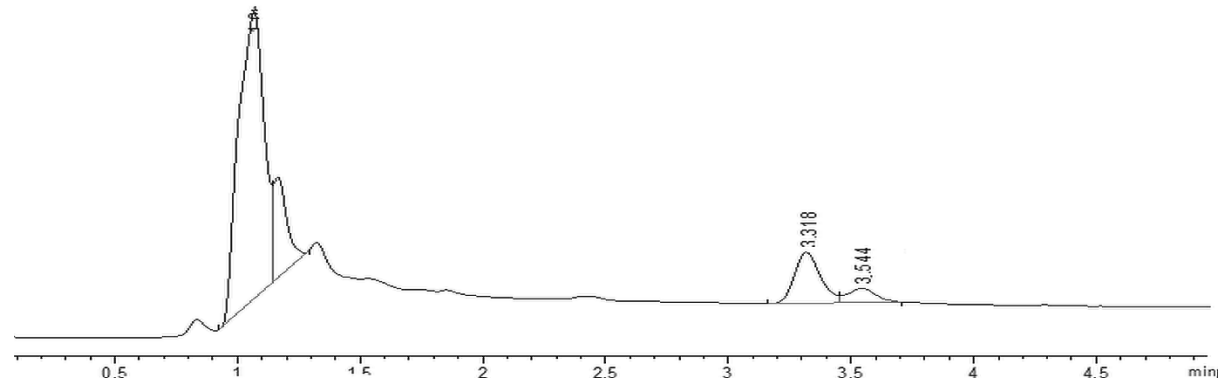
Weak acids, such as citric acid / malic acid interact with some alkaloids forming complexes.

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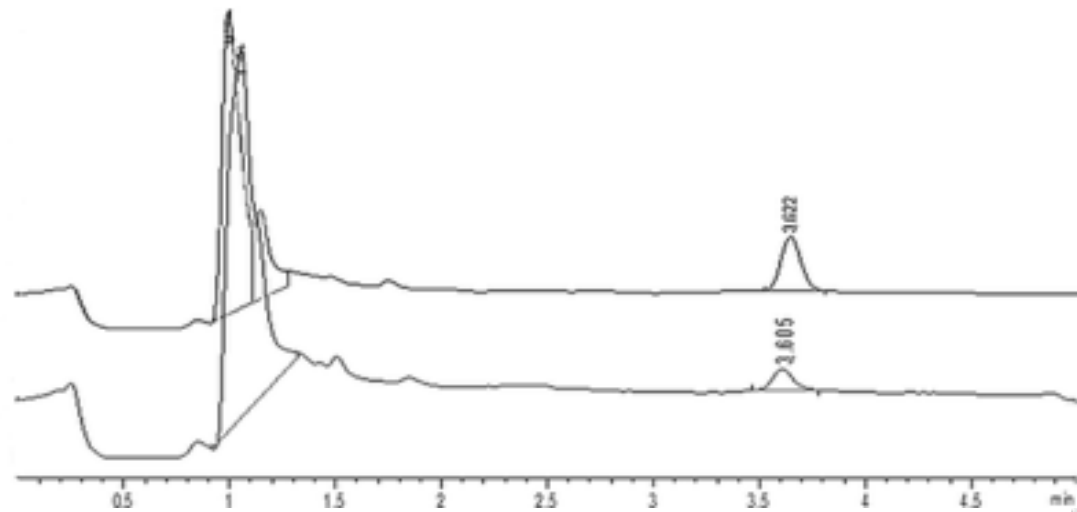


NADES extract of Narcissus (malic acid: glucose: water)

pH<10

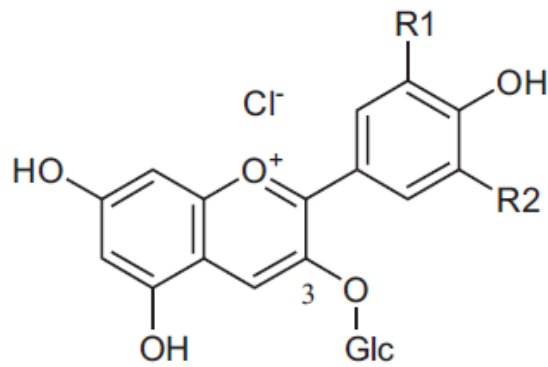


pH>10



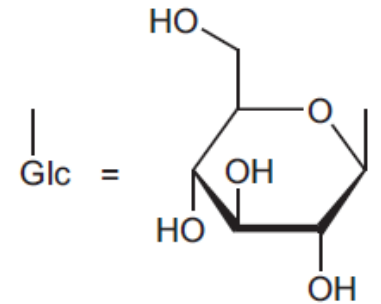
Example: Blueberry/ bilberry extract

Anthocyanins



Cyanidin	R1 = OH	R2 = H
Delphinidin	R1 = R2 = OH	
Malvidin	R1 = R2 = OCH ₃	
Peonidin	R1 = OCH ₃	R2 = H
Petunidin	R1 = OH	R2 = OCH ₃

3-monoglucosides



β -D-glucopyranosyl



HPLC/DAD- UV/Vis

(xylitol : citric acid: water); (glucose: citric acid : water)

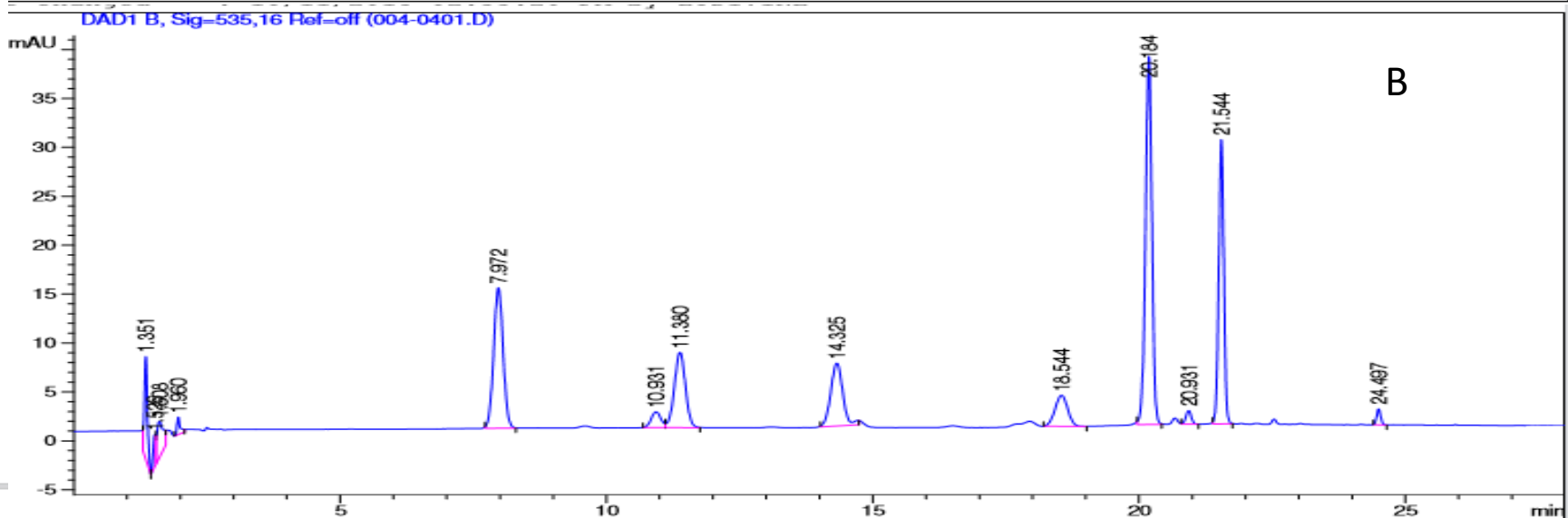
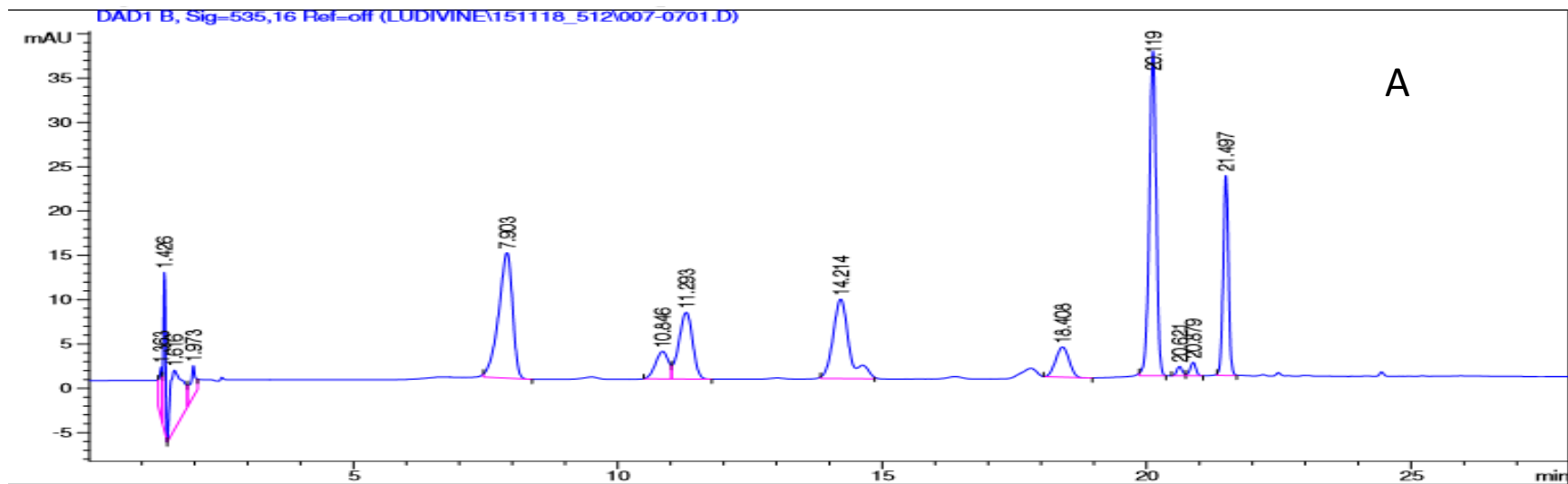
Possible with direct dilution or SPE

- Cannot be extracted with any immiscible organic solvent.
- Salting out/immiscible organic does not work.





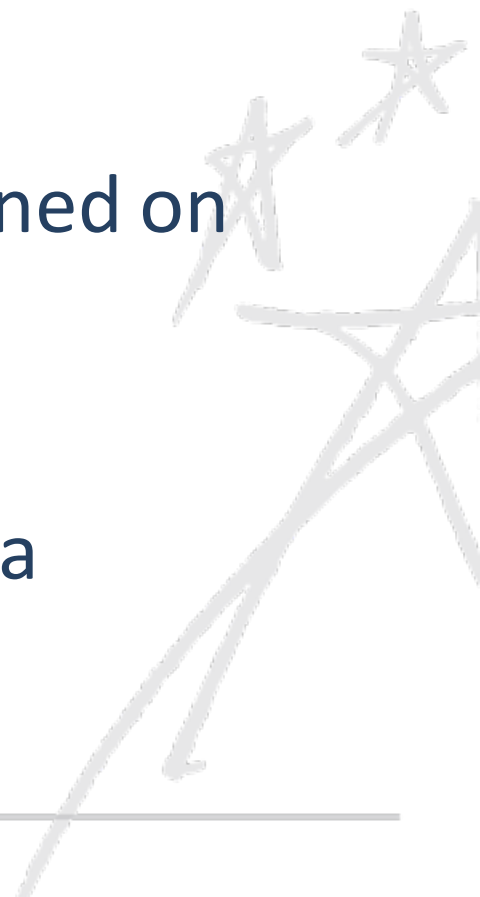
NADES blueberry (A) (EtOH:H₂O: HCl) blueberry (B)





LCMS with direct dilution?

- Possible and easy when analytes have higher R_t 's than NADES components:
 - NADES components: usually unretained on C18 column.
 - Divert the flow for first minute with a switching valve
-





Concluding....

Sample prep and analysis not a problem...but special consideration should be given to:

- **Extraction power** of hydrophilic polymers or large molecules with NADES is greater than usual solvents or even water.

Example: **lectins** in bulbs caused build-up of pressure in HPLC column (had to be removed).

Pectins are well extracted...not always welcome though!!



Universiteit Leiden

Thank you!!!

e.g.wilson@biology.leidenuniv.nl

