

Innovative Food Product Development Cycle: Frame for Stepping Up Research Excellence of FINS

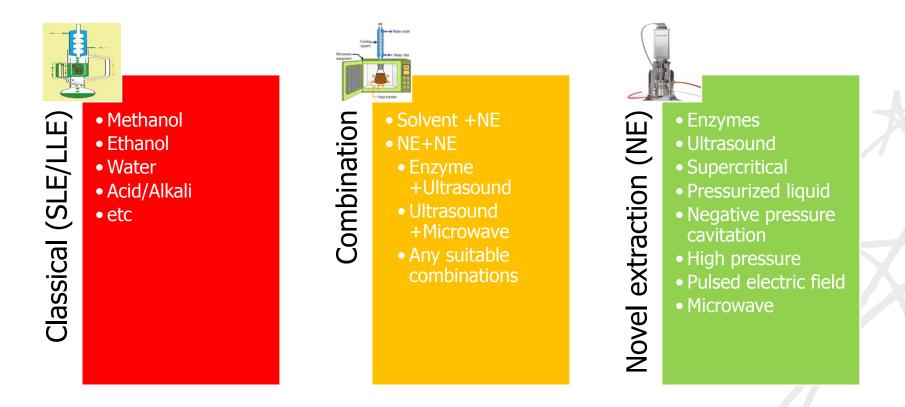


Conventional vs Novel extraction technologies

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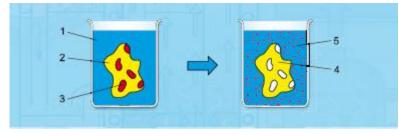
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Extraction technologies



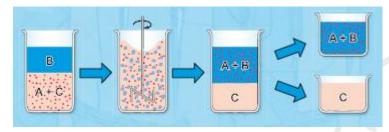
Conventional extraction techniques

Solid-liquid extraction



1 solvent, 2 extraction material (solid carrier phase with transition component), 3 transition component,
4 depleted solid carrier phase, 5 solvent with dissolved transition component

Liquid-liquid extraction



When the initial mixture (A+C) and the solvent (B) are mixed, the transition component (A) is transferred into the solvent.

After settling, two phases are obtained: the extract **(A+B)** and the carrier liquid **(C)**.

Selected example

Extractable Biomolecule	Substrate	Yield	
Pectin	Apple pomace, Citrus peel, Sugar beet, Sunflower heads, wastes from tropical fruits	10%–15%, 20%–30%	
Flavanones	Citrus peels and residues from segments and seeds after pressing		
Total and soluble dietary fibres	Apple pomace	72% and 10%	
Phenolic compounds	Apple pomace	33%	
γ-oryzanol	Rice bran	1527–4164 mg/kg	
β-glucans	Barley bran		
Lignins	Flaxseeds		
Phenolic acids	Wheat brans		
Lycopene and β-carotene	Tomato pomace	50%	

Example of some extracted bioactive compounds by different solvents

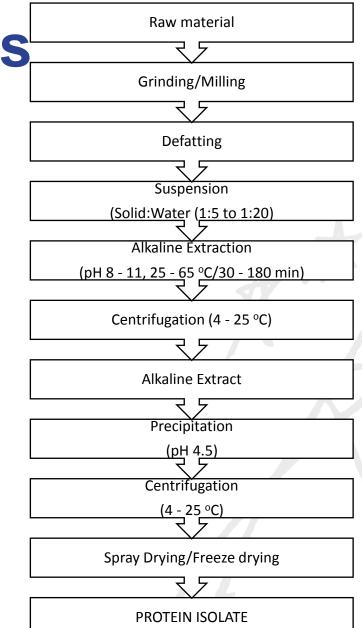
Water	Ethanol	Methanol	Chloroform	Dichloromethano I	Ether	Acetone
Anthocyanins	Tannins	Anthocyanin	Terpenoids	Terpenoids	Alkaloids	Flavonoids
Tannins	Polyphenols	Terpenoids	Flavonoids		Terpenoids	
Saponins	Flavonol	Saponins				
Terpenoids	Terpenoids	Tannins				
	Alkaloids	Flavones				
		Polyphenols				

Extraction of proteins

Technique employed for range of raw materials including grains, marine and meat sources

Challenge Removal of salt from final product





Drivers for novel extraction technologies in the food industry

- Regulation
- Food Safety and Shelf life extension
- Nutrient and Sensory aspects
- Consumer and Processor Acceptability
- Technology advances
- Clean and green extraction technologies Environmental impact
- Hurdle concept

Principle of clean and green extraction techniques

- **Principle 1:** Innovation by selection of varieties and use of renewable plant resources.
- Principle 2: Use of alternative solvents and principally water or agro-solvents.
- Principle 3: Reduce energy consumption by energy recovery and using innovative technologies.
- **Principle 4:** Production of co-products instead of waste to include the bio- and agro-refining industry.
- **Principle 5:** Reduce unit operations and favour safe, robust and controlled processes.
- **Principle 6:** Aim for a non denatured and biodegradable extract without contaminants.



Chemat (2012) Int. J. Mol. Sci., 13(7), 8615-8627

Novel extraction techniques

- Enzyme assisted extraction
- Microwave assisted extraction
- Ultrasound assisted extraction
- Supercritical fluid extraction
- Pressurized liquid extraction
- Negative pressure cavitation



Alternative solvents for green extraction

Solvent			Solvent Pov	wer	Health &		Environmental
	Extraction Technique (Application)		Weakly Polar	Non- Polar	Safety	Cost	Impact
Solvent-free	Microwave Hydrodiffusion and Gravity (antioxidants, essential oils)	+++	+		+++	+	+++
	Pulse Electric Field (antioxidants, pigments)	+++	+		+++	+	+++
	Steam distillation (essential oils)	++	+		+	++	+
Water	Microwave-assisted distillation (essential oils)	+++	+++	+	+	+	++
Water	Extraction by sub-critical water (Aromas)	+	++		+	+	+
CO ₂	Supercritical fluid extraction (decaffeination of tea and coffee)	-	+	+++	+	+	+
Ionic liquids	Ammonium salts (Artemisinin)	-	+	+++	-	-	++
	Ethanol (pigments and antioxidants)	+	+	-	-	++	+
Agrosolvents	Glycerol (polyphenols)	+	+	-	-	+	+
	Terpenes such as d-limonene (fats and oils)	-	-	++	-	+	+
Petrochemical							
solvents	n-Hexane (fats and oils)	-	+	+++		++	
<u>A</u> stars							

Enzyme assisted extraction

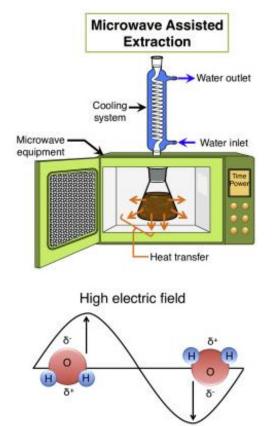
Application of enzyme assisted extraction for bioactives

Raw material	bioactive compound	enzymes used for extraction
Undaria pinnatifida	Fucoxanthin	Alginase lyase enzymes, temperature of 37 °C and pH of 6.2
Sargassum horneri	Antioxidant rich extracts	Carbohydrases and proteases
Brown seaweed species	Antioxidant rich extracts	Carbohydrases and proteases

Enzymes, pH and temperature employed for enzyme-assisted extraction of bioactives from seaweeds

Enzyme	temperature (°C)	рН	enzyme composition
Viscozyme	50	4.5	arabanase, cellulase, β -glucanase, hemi-cellulase and xylanase
Cellucast	50	4.5	group of enzymes catalyzing the breakdown of cellulose into glucose, cellobiose and higher glucose polymer
Termamyl	60	6.0	heat-stable α-amylase
Ultraflo	60	7.0	heat-stable multi-active β-glucanase
Neutrase	50	6.0	endoprotease
Flavourzyme	50	7.0	endoprotease and exopeptidase activities
Alcalase	50	8.0	α-endoprotease

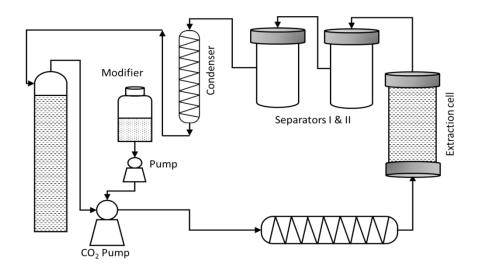
Microwave assisted extraction

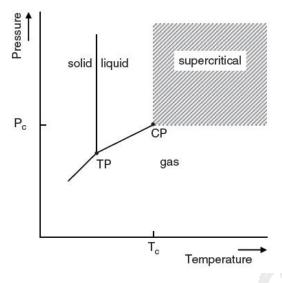


Molecular rotation and polarization

Marine algae	bioactive compound	conditions
Dunaliella tertiolecta	carotenoids	temperature of 56 °C and atmospheric pressure conditions
Fucus vesiculosus	fucoidan – sulphated polysaccharides	pressure of 200-800 kPa, extraction time 1–31 min, and alga/water ratio of 1/25 to 5/25 g ml 1
Porphyra (Nori) and Palmaria (Dulse), Undaria pinnatifida (Wakame), Himanthalia elongata (Sea spaghetti) and Laminaria ochroleuca (Kombu), Ulva Rigida (Sea Lettuce)	lodine	temperature of 200 °C, power of 1,000 W, holding time of 0-5 min

Supercritical fluid extraction

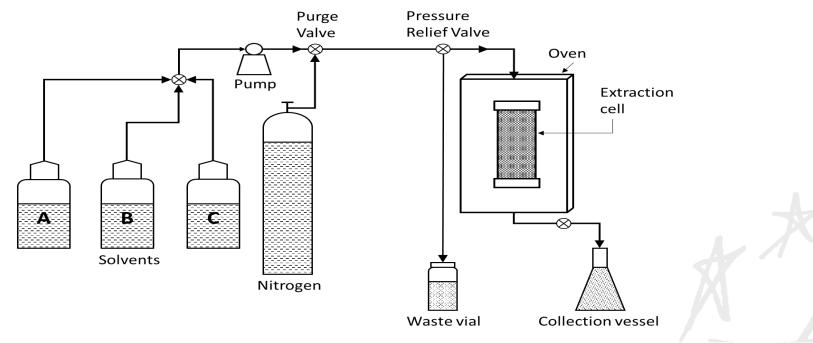




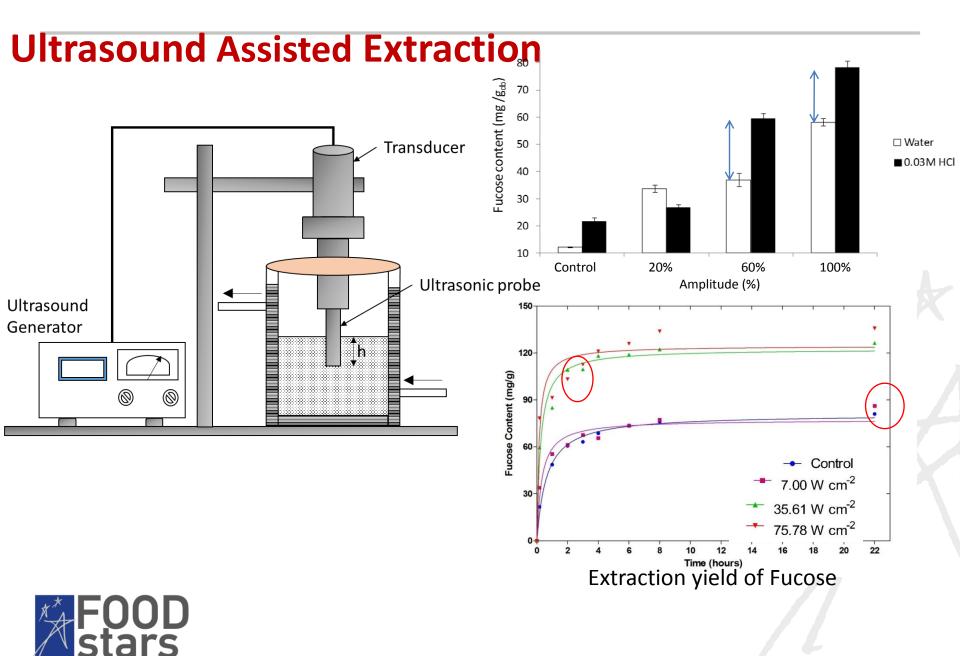
Marine algae	bioactive compound	conditions
Haemato coccuspluvialis	astaxanthin	ethanol along with acids were used as solvents for extraction
Scenedesmus almeriensis	carotenoids	pressure of 40 MPa and temperature of 60 °C
Dunaliella salina	chlorophyll	methanol as solvent
Hypneacharoides sp.	PUFA	temperature ranges from 40 to 50 °C and pressure from 24.1 and 37.9 MPa
Dunaliella salina	β-carotene	pressure of 30 MPa and temperature of 40 °C
Sargassum muticum	polyphenols	extractions were performed using CO ₂ modified with 12% ethanol at 15.2
		MPa pressure and 60 °C during 90 min



Pressurized liquid extraction

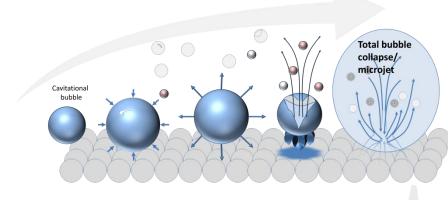


Marine algae	bioactive compound	conditions
Eisenia bicyclis	fucoxanthin	temperature 110 °C and 90% ethanol concentration
C. ellipsoidea	zeaxanthin	temperature and time for extraction were 115.4 °C and 23.3 min
Dunaliella salina	bioactive phenols	temperature of 40, 100 and 160 °C and time of 5, 17.5 and 30 min
Himanthalia elongate	bioactive phenols	temperature of 50, 100, 150 and 200 °C for 20 min
Undaria pinnatifida	antioxidants	water as solvent
Sargassum muticum	polyphenols	pressure of 10.3 MPa at 120 °C temperature for 6 min

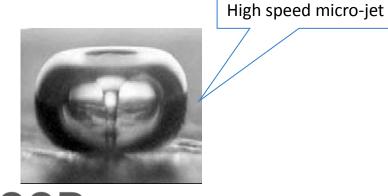


Mechanisms of action

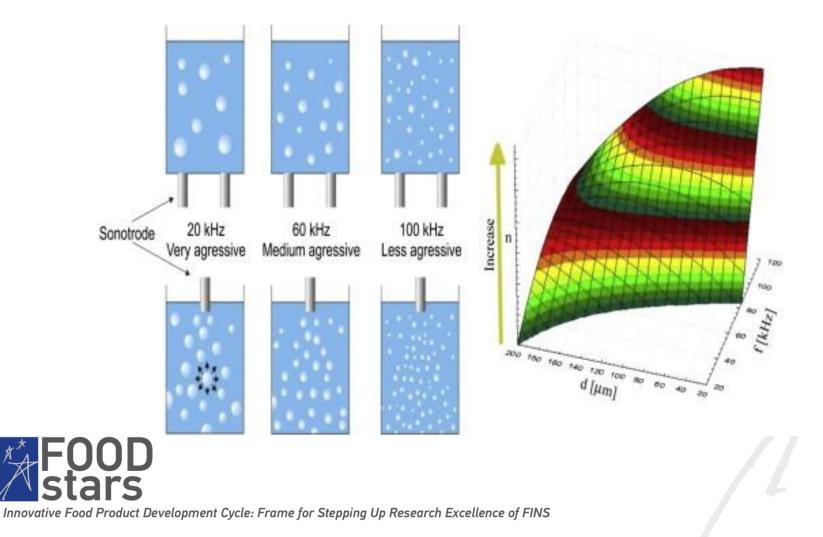


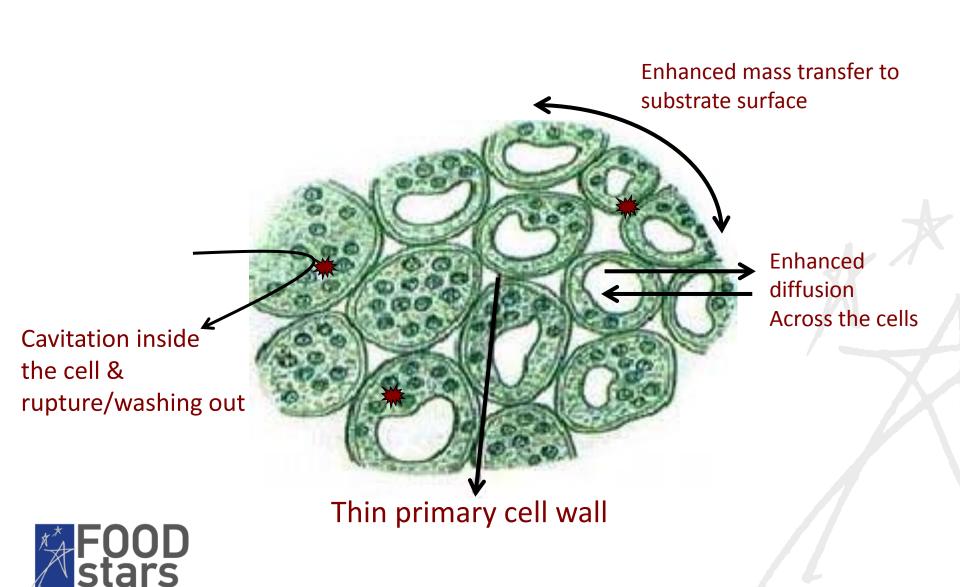


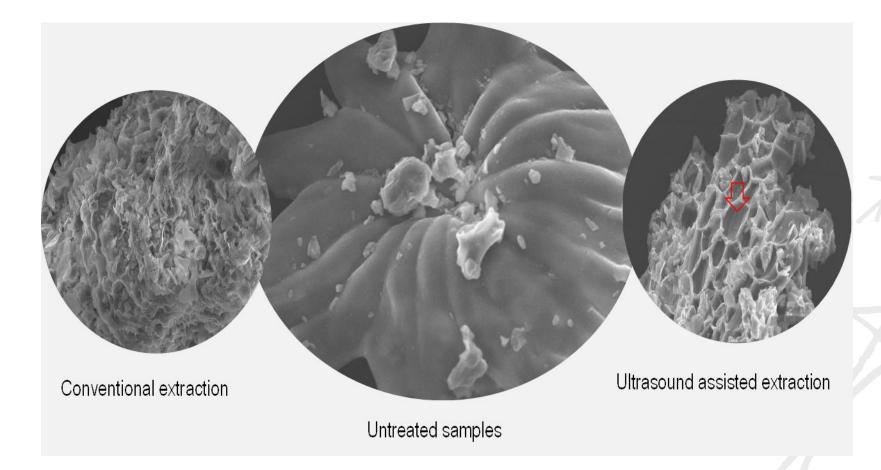


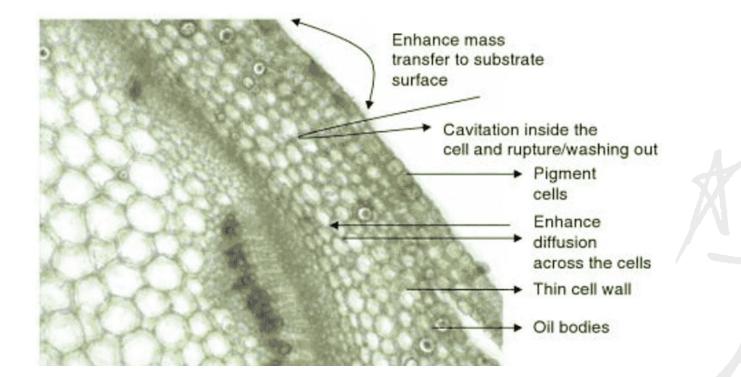


Size of the cavitational bubbles in dependence of ultrasounds frequency

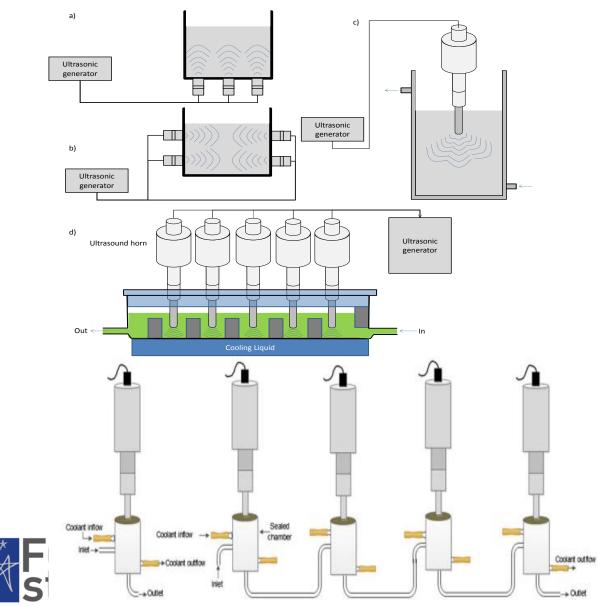








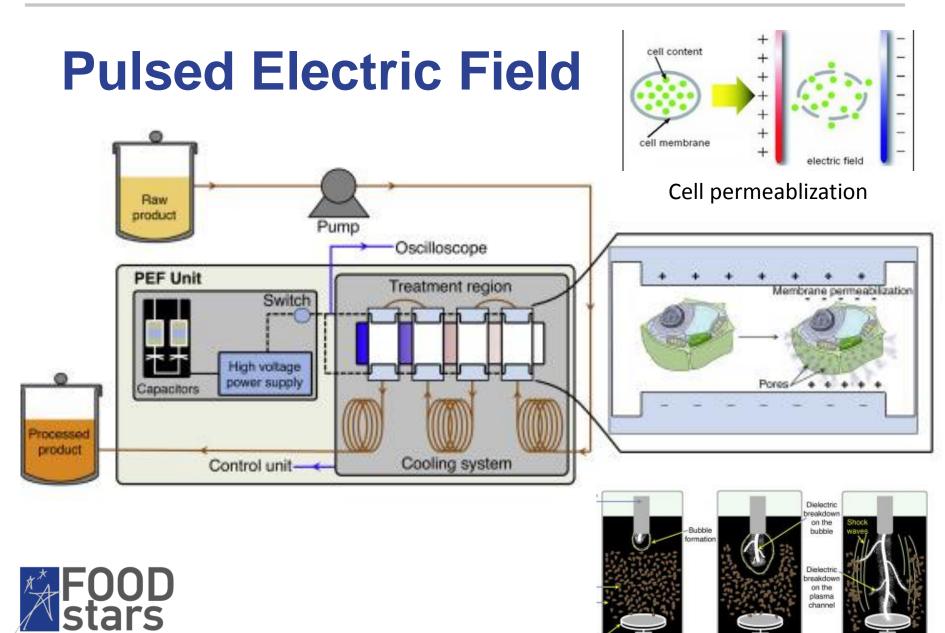
Commercially available ultrasonic systems

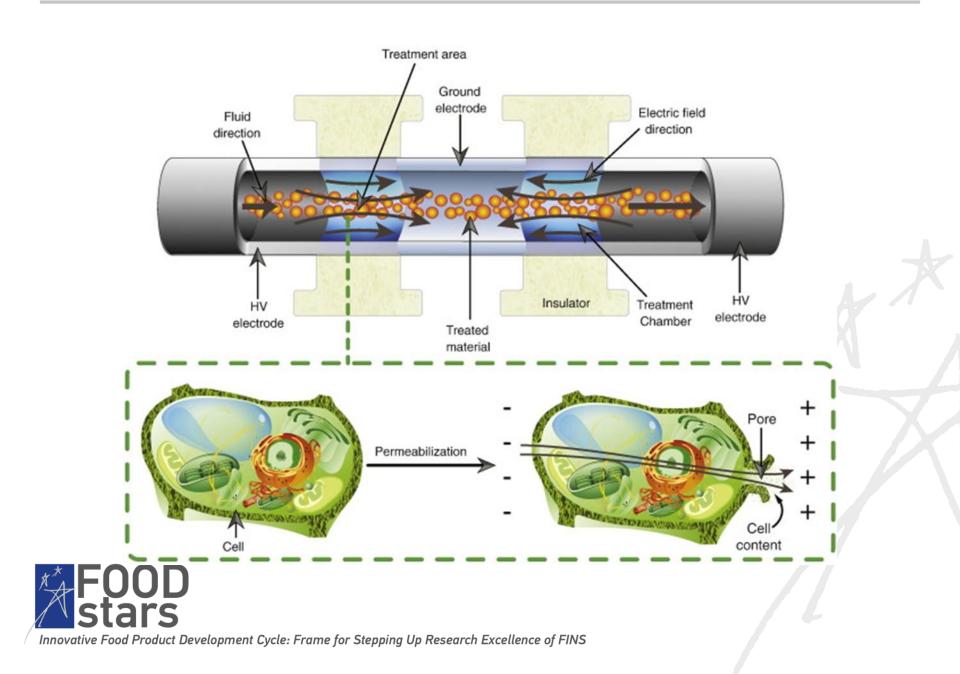


Ultrasound probe type system

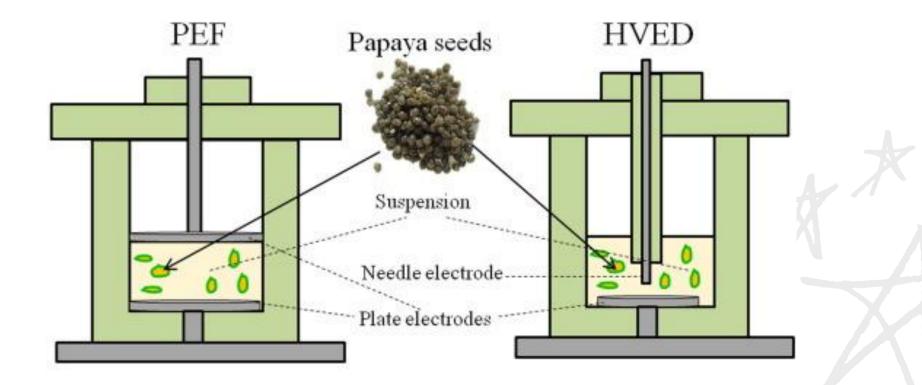


Ultrasonic bath system

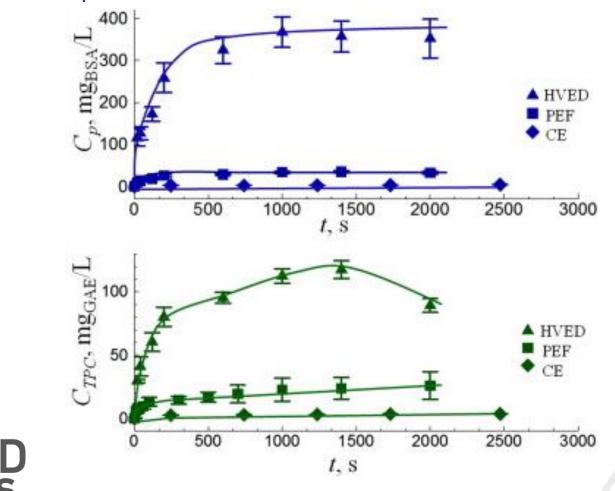




PEF and HVED treatment chambers



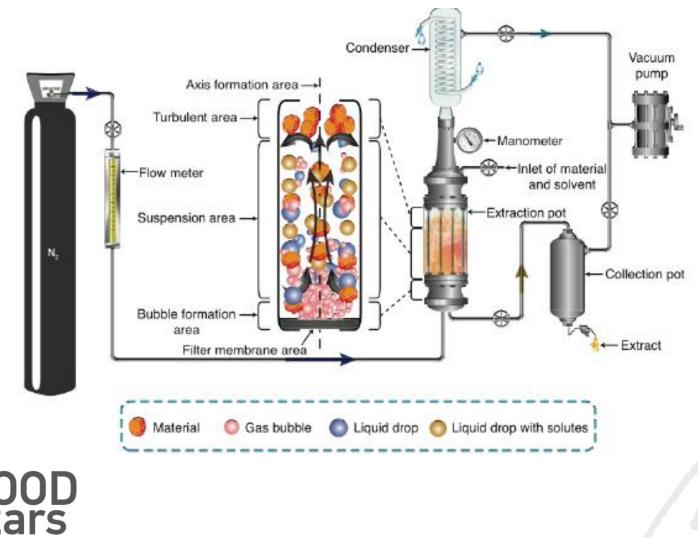
Concentration of proteins, and total phenolic compounds versus time of extraction, *t*, pulsed electric fields (PEF) and high voltage electrical discharges (HVED)-assisted extraction and conventional extraction (CE) at T = 20 °C and pH = 7

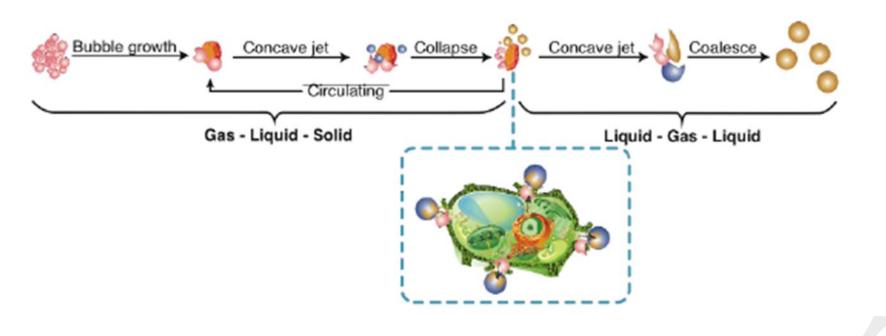


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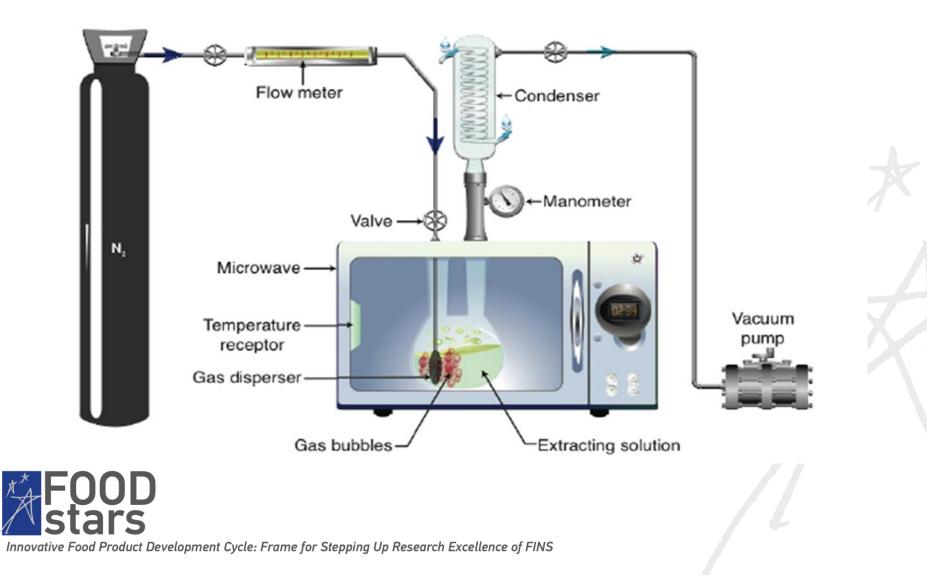
Parniakov et al., (2015)

Negative pressure cavitation





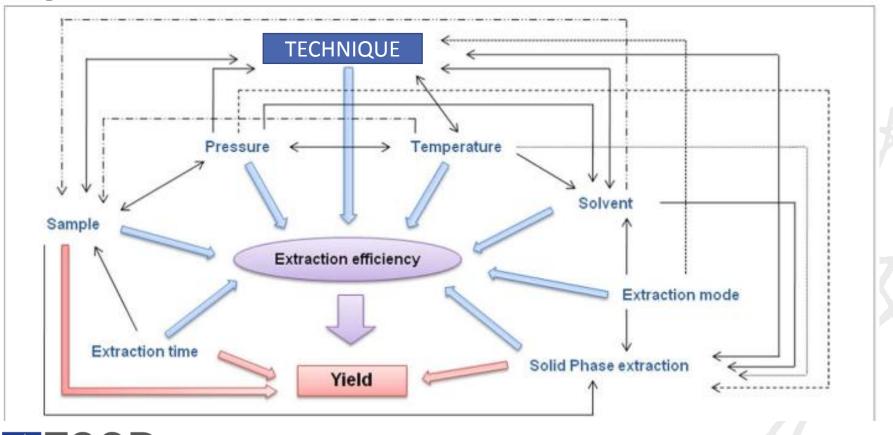




Combination techniques

Combination	Extraction conditions	Matrix and target compound
Supercritical fluid	Ultrasonic power: 180 – 360 W; extraction time (60 – 240 min);	Capsaicinoids and
extraction and	Ultrasonic frequency of 20 kHz	phenolics from
ultrasound	SFE: 15 \pm 0.5 MPa and temperature of 40 \pm 3 °C, CO ₂	malagueta pepper
Supercritical fluid	SFE: temperature (40, 50 and 60 °C), pressure (15, 20 and	Antioxidants and
extraction and	25 MPa), and ultrasound power (0, 200 and 400 W).	anthocyanins from
ultrasound	Ethanol and water as co-solvent	the blackberry
		bagasse
Supercritical fluid	15 min ultrasound-assisted extraction (static extraction)	Oleanolic acid and
extraction and	followed by SC–CO ₂ extraction, water content in ethanol	Ursolic acid
ultrasound	modifier (60–100%)	from Scutellaria
		barbata
Microwave and	Ultrasonic probe made of polyether ether ketone (frequency	Oil from soybean
Ultrasound	25 kHz; power 60W); Microwave 100 W)	germ and Seaweeds
	Temperature 45°C; extraction time 1 h	
Microwave and	microwave power 98W; Microwaves delivered to ultrasonic	Lycopene from
Ultrasound	bath operating at 40 kHz and power of 50 W	tomato paste
High pressure and	Ultrasonic bath 47 kHz ultrasonic extraction experiments	Soluble matter from
Ultrasound	assisted and non-assisted by pulsed hydrostatic pressure	mate leaves

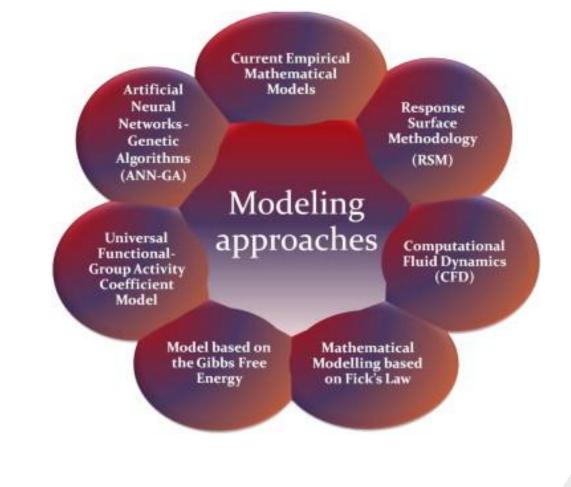
Factors affecting extraction yield



Factors affecting extraction yields

External factors	Key facts	
Extraction temperature	High temperature aid in disruption of interaction of solvent and matrix	
	High temperature enhances solvent diffusion rates	
	Low temperature enhances cavitation	
	Low temperature reduces thermal impact on target compound	
Extraction time	Long extraction time enhances extraction yields	
	Long extraction time may induce undesirable changes in the extracted compound.	
Solvent properties	Viscous solvent reduces diffusion	
	Volatile solvent may evaporate if extraction is carried out at higher temperature for long	
	duration	
	Polarity and solubility of target compound in the solvent	
Matrix	Particle size	
	Solvent matrix interaction	
	Ratio of solvent to matrix	

Modelling tools



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Conclusions

- ✓ Clear recent scientific advances in highlighting the potential applications of novel extraction techniques.
- Novel techniques will be required to address key emerging challenges faced by the food industry.
- ✓ Further research required to facilitate industry adoption of recent advances while convincing end users.



DISCLAIMER:

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the opinion of authors and not the opinion of European Commission.



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